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Registry No.—4b, 67382-63-2; 4c, 67382-64-3; 5b, 67382-65-4; 5c, 54781-19-0; 7b, 67382-66-5; 7c, 67382-67-6; 8b, 67382-68-7; 8c, 6651-36-1; 12, 67382-69-8; 16, 3839-48-3; triethylsilyl phenyl ether, 5888-66-4; tetrachloroethylene, 127-18-4.

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Solvolvsis of Virescenol B 19-Tosvlate

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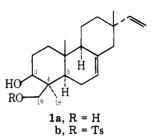
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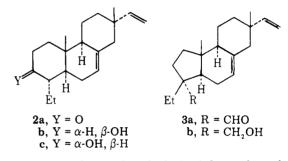
In continuation of a study of the chemistry of virescenol B (1a)¹ the aglycon of several of the fungal, virescenoside metabolites,³ the solvolysis of the 19-tosylate $(1b)^4$ in dimethyl sulfoxide was investigated. It produced two carbonyl-containing substances whose structures are the subject of this note.

One of the products could be shown to be ketone 2a on the basis of the following facts. Its infrared absorption at 1708 cm^{-1} revealed it to be a cyclohexanone. The disappearance of the oxymethine, oxymethylene, and 4-methyl ¹H NMR signals normally associated with the C(3) and C(4) substitution pattern of the virescenol B skeleton and the exhibition of a methyl triplet (J = 6 Hz) in the ¹H NMR spectrum of the product suggested the latter to be an α -ethyl ketone. Finally,

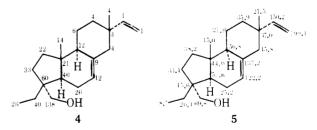
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its ¹³C NMR spectra confirmed the structural changes at C(3) and C(4), showed rings B and C of virescenol B $(1a)^5$ to be affected only minimally, and ring A reminiscent of a 3-ketosteroid.⁶ Sodium borohydride reduction of the ketone yielded a ca. 2:1 mixture of alcohols, whose ¹H NMR spectra indicated them to be equatorial and axial isomers, respectively. The low equatorial-axial isomer ratio, in contrast to the high ratio resulting from the hydride reduction of a 3-ketosteroid,⁷ was in accord with the presence of a 4α -ethyl-3-keto system whose ethyl group offered resistance to the normal α attack by borohydride. The ¹³C NMR spectra of the alcohols confirmed fully structures 2b and 2c for the reduction products.



The minor product of the solvolysis of 1b was shown by its infrared absorption bands of 2670 and 1722 cm^{-1} to be an aldehyde and by its ¹H NMR spectrum to have its carboxaldehyde unit in an equatorial orientation⁸ next to a nonprotonated carbon center. Once again the structural change at C(3) and C(4) was revealed not only by the new carbonyl group, but also by a methyl triplet (J = 5 Hz) indicative of the presence of an ethyl group. Sodium borohydride reduction of the aldehyde yielded an alcohol whose ¹H NMR spectral characteristics revealed the presence of an equatorial hydroxymethyl group^{8,9} next to a nonprotonated carbon and an ethyl group. The ¹³C NMR spectra of the alcohol exhibited virescenol B-like ring B and C carbon signals and a homoneopentyl carbon signal customary for a methyl group on an ethyl function terminating on a nonprotonated carbon site. A Yb(DPM)₃ shift study (cf. $\Delta\delta$ values on formula 4) permit-

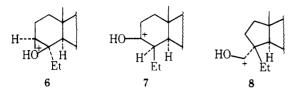


ted a carbon signal assignment and structure analysis as depicted by formula 5 (3b), thus showing the aldehyde to possess structure 3a.

The simplest explanation for the production of the two carbonyl compounds, 2a and 3a, on solvolysis of tosylate 1b involves the migration of the 4α -methyl group to the site of the departing tosylate group with concommitant O-C(4) bond formation by the neighboring 3β -hydroxy group. Hydride migration from C(3) to C(4) of the resultant conjugate acid of a 3β , 4β -epoxide (6) leads to an O-protonated 4β -ethyl 3-

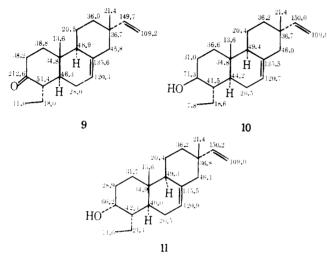
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Notes



ketone (7), whose enolization and reketonization gives ketone 2a, whereas C(2)-C(3) bond migration to C(4) of 6 yields the conjugate acid (8) of aldehyde 3a.

The carbon shifts of ketone 2a are outlined on formula 9. The similarity of the C(2) shift of 38.2 ppm with that of 5α androstan-3-one (38.1 ppm)⁶ shows the ethyl group to be equatorial and hence 4α oriented. The assignment of the chemical shifts of the alcohols 2b and 2c is portrayed on formulas 10 and 11, respectively.



The migration of C(18) from C(4) to C(19) in the solvolysis of virescenol B 19-tosylate (1b) may be of biogenetic significance. Whereas many one-carbon rearrangements abound in the pimaranic diterpene field, none of the aforementioned type has been observed heretofore. Nevertheless, an ethano unit attachment to C(4) of diterpene rings A, such as in ketone 2a, appears in the form of a furan ring in some constituents of the coffee bean.¹⁰

Experimental Section

Melting points were determined on a Reichert micro-hotstage and are uncorrected. Infrared spectra were obtained on a Perkin-Elmer 167 spectrophotometer. ¹H NMR spectra (Me₄Si, $\delta = 0$) were recorded on a Jeol H-60 spectrometer and the ¹³C NMR spectra were produced on a Varian XL-100-15 spectrometer operating at 25.2 MHz in the Fourier transform mode.

Solvolysis of Virescenol B Tosylate (1b). A solution of 600 mg of tosylate 1b in 7 mL of dimethyl sulfoxide was heated at 95 °C under nitrogen with stirring for 7 h. After the addition of 50 mL of saturated brine solution the mixture was extracted thoroughly with chloroform. The extract was washed with water, dried (Na_2SO_4) , and evaporated under vacuum. Chromatography of the residual oil, 330 mg, on silica gel and elution with benzene yielded 60 mg (16%) of liquid aldehyde **3a:** IR (CCl₄) 2670 (CHO), 1722 (C=O) cm⁻¹; NMR (C₆D₆) δ 0.68, 0.86 $(s, 3 \text{ each}, Me_2), 0.73 (t, 3, J = 6 \text{ Hz}, Me \text{ of Et}), 8.96 (s, 1, CHO)$

Anal. Calcd for C₂₀H₃₀O: C, 83.86; H, 10.56. Found: C, 83.70; H, 10.72

Elution with 20:1 benzene–ethyl acetate gave 185 mg (50%) of solid whose crystallization from pentane led to crystalline ketone 2a: mp 92-94 °C; IR (CCl₄) 1708 cm⁻¹ (C=O); NMR (C₆D₆) δ 0.76, 0.90 (s, 3 each, Me_2), 0.96 (t, 3, J = 6 Hz, Me of Et).

Anal. Calcd for C₂₀H₃₀O: C, 83.86; H, 10.56. Found: C, 83.92; H, 10.54.

Alcohols 2b and 2c. A solution of 60 mg of sodium borohydride in 5 mL of ethanol was added dropwise to a stirring solution of 600 mg of ketone 2a in 15 mL of ethanol at 0 °C. The mixture then was stirred at room temperature for 1 h and poured into ice water. It was extracted with chloroform and the extract dried (Na₂SO₄) and evaporated. Chromatography of the oily residue, 550 mg, on silica gel and elution with 20:1 benzene-ethyl acetate afforded 180 mg (30%) of semisolid alcohol 2c: IR (CCl₄) 3625 cm⁻¹(OH); NMR (CCl₄) § 3.86 (m, 1 OCH).

Anal. Calcd for C₂₀H₃₂O: C, 83.27; H, 11.18. Found: C, 83.42; H, 11.02.

Further elution with the same solvent pair gave 310 mg (51%) of solid whose crystallization from pentane-benzene yielded crystalline alcohol 2b: mp 120-125 °C; IR (CCl₄) 3620 cm⁻¹(OH); NMR (CCl₄) δ 3.20 (m, 1, OCH).

Anal. Calcd. for C₂₀H₃₂O: C, 83.27; H, 11.18. Found: C, 83.38; H, 11.00.

Alcohol 3b. A solution of 15 mg of sodium borohydride in 5 mL of ethanol was added dropwise to a stirring solution of 50 mg of aldehyde 3a in 3 mL of ethanol at 0 °C. The mixture then was stirred at room temperature for 1 h. Workup as above, chromatography of the curde product, 40 mg, on silica gel, and elution with 30:1 benzene-ethyl acetate led to 35 mg (80%) of semisolid alcohol 3b: NMR δ 3.25, 3.45 $(AB dd, 2, J = 11 Hz, OCH_2)$.

Anal. Calcd for C₂₀H₃₂O: C, 83.27; H, 11.18. Found: C, 83.05; H, 11.31.

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Novel Applications of the Potassium Chlorate-Osmium Tetroxide Oxidizing System. Synthesis of α -Dicarbonyl Derivatives from Acetylenic Compounds. Synthesis of a 2,3-Dihydroxy-1,4-dione from a 2,5-Dialkylfuran

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The cis hydroxylation of olefins to α -diols by metal chlorates in aqueous solution containing catalytic amounts of osmium tetroxide has found wide use.¹ This method is especially useful when the oxidation of other organic functions present in the substrate has to be avoided; most of these functions are, in fact, unaffected by the said oxidizing system. The reaction of acetylenic compounds with this oxidizing